

# Appendix 1

## Method and results of Pollen and NPP analysis.

### Sampling

A sample was taken from a patch of straw, presumably used as packing material in hanging vessel figure 3a. The sample was treated for with a standard pollen preparation procedure following Fægri and Iversen 1989.

A soil/dirt sample was taken from the inside of the hanging vessel figure 3d during excavation.

### Preparation procedure:

The sample was subjected to a relatively gentle preparation procedure developed to yield the highest frequency and diversity of non-pollen palynomorphs (Enevold 2018, 2019).

#### 0. Before chemical preparation

- a) Weigh 2-3 g of sample.
- b) Addition of 1-3 *Lycopodium* tablets.

#### 1. Disaggregation

- a) Add HCl (10%) until reaction stops and calcium is dissolved.
- b) Add demineralised and  $\mu$ -filtered water until 10ml.
- c) Centrifuge for 7 min., 4200-4500rpm.
- d) Decant and wash three times.
- e) Add 10 ml KOH (10%) and heat in water bath for 10-15 min.
- f) Cool and wash twice.

#### 2. Filtering

- a) Use 212  $\mu$ m filter with demineralised and  $\mu$ -filtered water.
- b) Use 6  $\mu$ m filter with demineralised and  $\mu$ -filtered water.
- c) Centrifuge for 5 min., 4500rpm.
- d) Decant and wash one-three times.

#### 3. Separation process

- a) Add 3 ml Sodium polytungstate SPT (specific gravity of 1.8-2 g/cm<sup>3</sup>).
- b) Whirlimix for approx. 30 seconds.
- c) Centrifuge for 15 min., 2500rpm.
- d) Separate sample from residue.
- e) Wash two-three times.

#### 4. Addition to medium

- a) Wash once in 96 % ethanol.
- b) Wash once in 99.9 % ethanol.
- c) Add 2-3 ml tertiary butanol and 3-7 drops of silicon oil.
- d) Dehydrate at 50 grader for 24-48 hours.

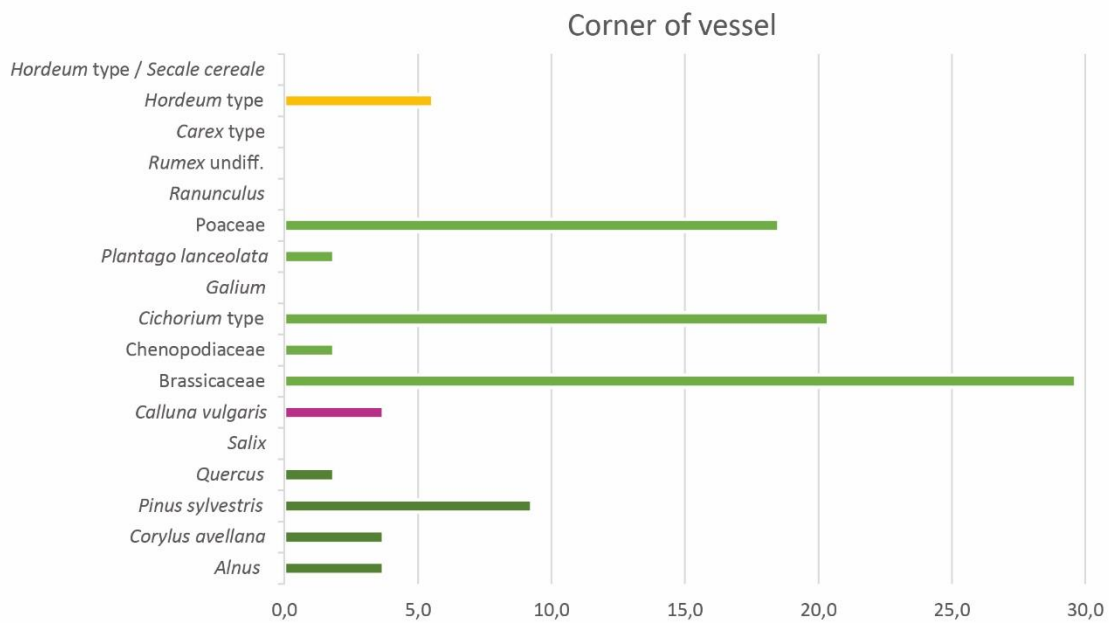
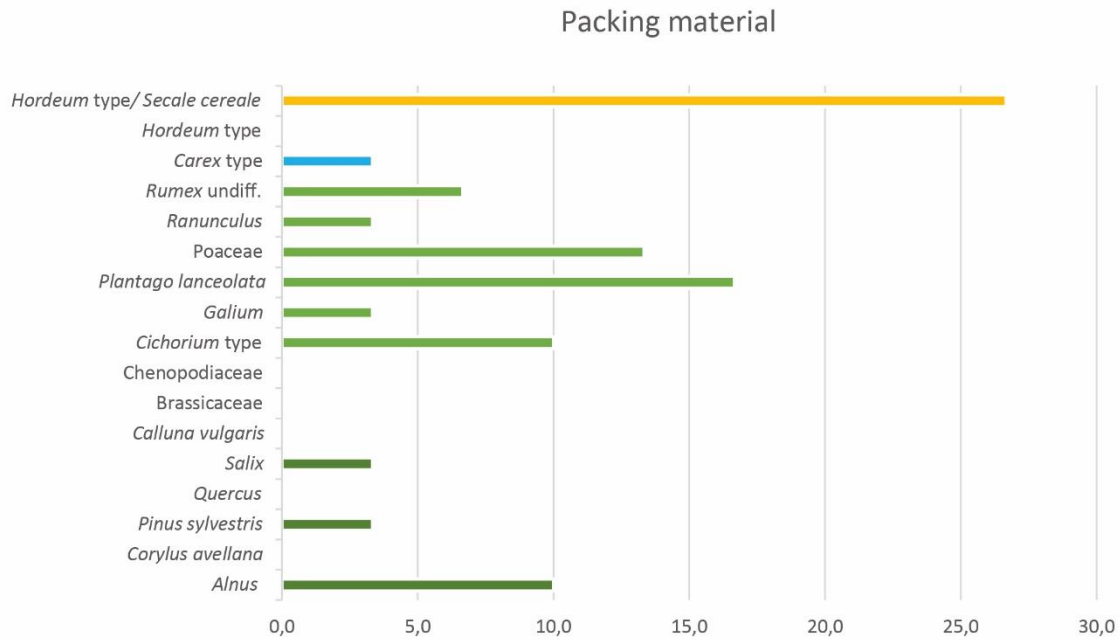


Figure 1: Histogram showing all recovered pollen types (y-axis) as percentages (x-axis) in a sample from the packing material in the hanging vessel figure 3a and from a corner of the hanging vessel figure 3d.

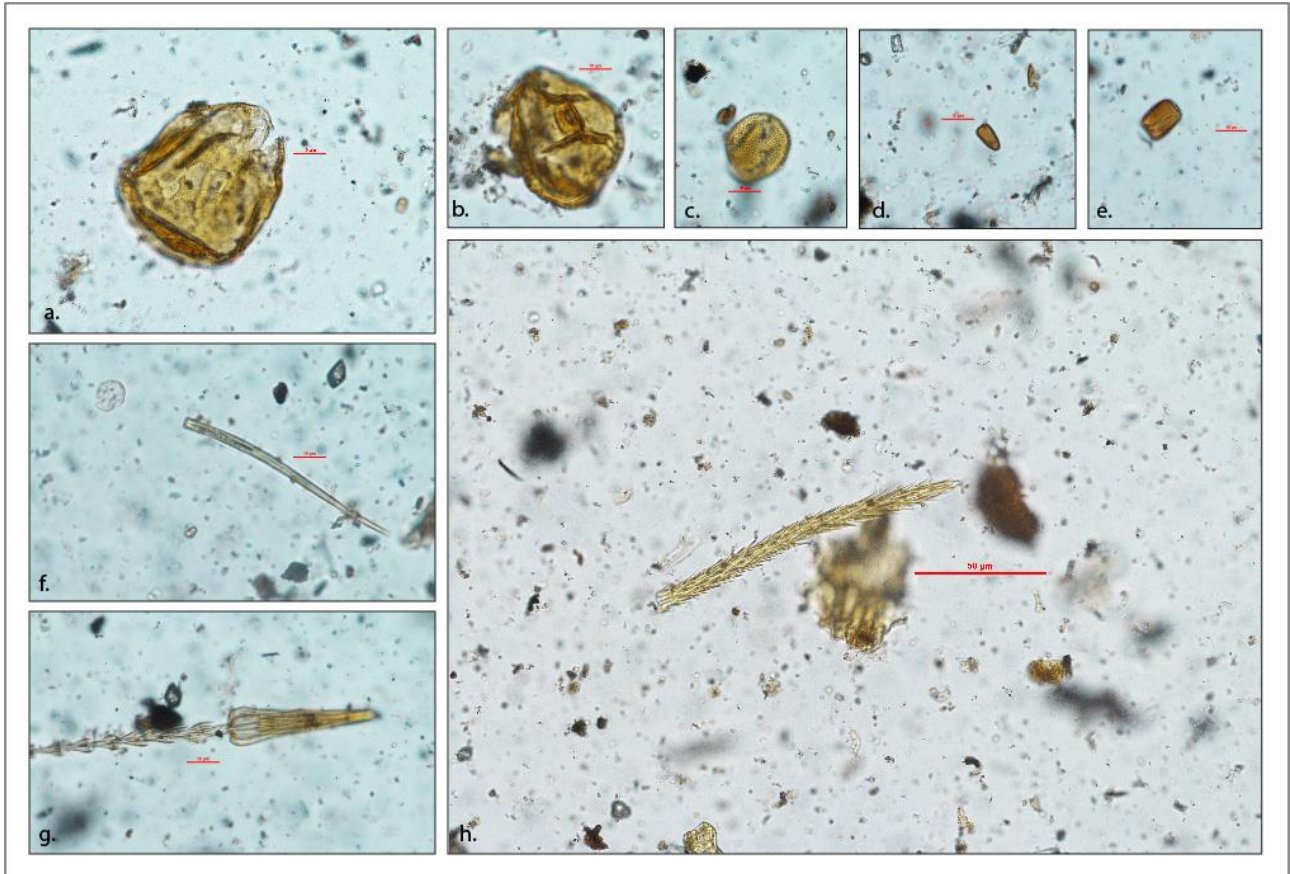


Figure 2: Microscope photos of selected palynomorphs found in the samples: a. *Hordeum* type pollen, b. *Cerealia* undifferentiated, c. *Brassicaceae* pollen, d. *Sporormiella* spore (small), e. *Sporormiella* spore (large), f. bee hair, g. unidentified and h. insect antenna. Photos: Renée Enevold.

Table 1 All non-pollen palynomorphs recorded in the sample from the vessel. Where possible, names have been given to the NPP type according to previous given number in a collection (MM = Moesgaard Museum, HdV = Hugo de Vries laboratory).

Number	Identification	P3	%
MM - 560	Plant fragment	4	1.6
NPP - 7	Plant fragment	1	0.4
MM - 487	Plant fragment	3	1.2
MM - 457	Small-pitted perforation plate	3	1.2
<b>Plant fragments total</b>		<b>11</b>	<b>4.5</b>
MM - 827	Insect antenna	23	9.4
NPP - 9	Insect hair	3	1.2
<b>Insect fragments total</b>		<b>26</b>	<b>10.6</b>
NPP - 3	Fungal spore	6	2.4
NPP - 6	Fungal hyphae	3	1.2
MM - 231	Fungal hyphae	1	0.4
MM - 328	Fungal spore	1	0.4
MM - 154	Fungal spore	3	1.2
MM - 334	Fungal spore	1	0.4
MM - 65	Fungal spore	14	5.7
MM - 434	Fungal spore	2	0.8
MM - 158	Fungal spore	11	4.5
MM - 293	cf. <i>Schitzothecium</i>	2	0.8
<b>Fungal remains total</b>		<b>44</b>	<b>18.0</b>
HdV - 7A	<i>Chaetomium</i> sp.	16	6.5
MM - 220 (HdV - 113)	<i>Sporormiella</i> small type	18	7.3
MM - 269	<i>Cercophora</i> type	1	0.4
MM - 32 (HdV - 113)	<i>Sporormiella</i> large type	1	0.4
HdV - 55A	<i>Sordaria</i> type	2	0.8
<b>Spores from coprophilous fungi total</b>		<b>38</b>	<b>15.5</b>
Charcoal	Charcoal fragments >20 µm	74	30.2
<b>Charcoal total</b>		<b>74</b>	<b>30.2</b>
Hdv - 62	Zygnemataceae	1	0.4
<b>Algae total</b>		<b>1</b>	<b>0.4</b>
NPP - 4	Unidentified	38	15.5
NPP - 5	Unidentified	13	5.3
<b>Unidentified total</b>		<b>51</b>	<b>20.8</b>
Sum		245	100.0

## Bibliography

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